Reprint

International Journal of Sustainable Crop Production (IJSCP)

(Int. J. Sustain. Crop Prod.)

Volume: 11

Issue: 3

August 2016

Int. J. Sustain. Crop Prod. 11(3): 8-11 (August 2016)

EFFICACY OF INDIGENOUS Trichoderma STRAIN ON FOOT ROT OF TOMATO SEEDLINGS INDUCED BY Rhizoctonia solani UNDER GREENHOUSE CONDITIONS

M.M. ISLAM, A. AKTER AND M.D. HOSSAIN



EFFICACY OF INDIGENOUS Trichoderma STRAIN ON FOOT ROT OF TOMATO SEEDLINGS INDUCED BY Rhizoctonia solani UNDER GREENHOUSE CONDITIONS

M.M. ISLAM^{1*}, A. AKTER² AND M.D. HOSSAIN³

¹Department of Plant Pathology and Seed Science, Sylhet Agricultural University, Sylhet; ²Department of Agricultural Extension Education, Sylhet Agricultural University, Sylhet; ³Department of Plant Pathology, Bangladesh Agricultural University, Mymensingh.

^{*}Corresponding author & address: Md. Monirul Islam, E-mail: mmislambau@gmail.com Accepted for publication on 18 July 2016

ABSTRACT

Islam MM, Akter A, Hossain MD (2016) Efficacy of indigenous *Trichoderma* strain on foot rot of tomato seedlings induced by *Rhizoctonia* solani under greenhouse conditions. *Int. J. Sustain. Crop Prod.* 11(3), 8-11.

Tomato (*Solanum lycopersicum*) is one of the important crops among all the vegetables and very popular vegetable in Bangladesh. As the pathogen, *R. solani* is a soil borne pathogen and prefers damp, especially water logged conditions, the incidence of foot rot has been found more and causes a great economic loss every year in all tomato growing areas. We evaluated three *Trichoderma* isolates (*T. harzianum* TR05, *T. virens* TR06 and *T. asperellum* TR08) originating from Bangladesh as potential biological control agents against foot rot of tomato under greenhouse conditions. After seed treatment with TR05, there was a lower disease incidence (5.79%) of foot rot of tomato seedlings than for the other isolates, and the germination percentage of the tomato seedlings was also highest (89.7%). The other growth parameters were also promoted by inoculation with the *Trichoderma* isolates; particularly TR05 showed statistically higher root length (13.5 cm), shoot length (12.7 cm), dry weight (5.40 g) and seedlings.

Key words: trichoderma, rhizoctonia solani, tomato, foot rot, seed treatment

INTRODUCTION

The pathogen, Rhizoctonia solani is occurs in tropical and sub-tropical zone of the world where high temperature prevails. The pathogen attacks economically important plants including vegetables, ornamental, cereals, field crops and weeds. Some of the common hosts include legumes, crucifers, tomato, pulse, peanuts and tobacco in which the pathogen causes foot rot or root rot (Anahosur 2001). The infected plant shows rot of tissue, dark sunken lesions and girdle the stems adjacent to soil level termed as collar region causing death by disrupting the translocation of food from top to root as well as by breaking at the point of infection. As a result, the pathogen causes a great economic loss in various crops. As the pathogen R. solani, is a ubiquitous, cosmopolitan saprophyte & being poly phages, it appears every year in all tomato growing regions of Bangladesh. Because of *R. solani* is chiefly a seed-bed disease; young and tender plants infected in the seed bed and carry the disease to the field. As a result, the plants may be attacked at any age. Under favorable conditions of the disease, it may outbreak in epiphytotic forms. As a result, it affects yield and quality or cause total failure of the crop incurring substantial losses to the economy of the country. Moreover, Tomato (Solanum lycopersicum) is one of the important crop among all the vegetables and very popular vegetable crop of Bangladesh. As the pathogen, R. solani is a soil borne pathogen and prefers damp, especially water logged conditions (Fakir 2000), the low laying areas during summer season of Bangladesh, the incidence of foot rot has been found more (Babar 1999). Therefore, control of this disease is very important. Various methods have been investigated for controlling R. solani, including chemical controls, cultural practices, resistant variety use, plant extracts, plant volatile compounds and biological control. Although chemical control is an important method for eradication of the pathogen in severe attack, it is not economical in the long term because it pollutes the atmosphere, damages the environment, leaves harmful residues and can lead to development of resistant strains among the target organisms with repeated use (Naseby et al. 2000). Biological control is an alternative to control the pathogen at low cost and to restore soil fertility without disturbing other components of the environment.

Among the hundreds of organisms identified as potential biocontrol agents, only a few have been commercially acceptable to control *R. solani* (Warrior *et al.* 2002). In our previous study, we obtained three *Trichoderma* isolates, *T. harzianum* TR05, *T. virens* TR06 and *T. asperellum* TR08, from different locations in Bangladesh, which have potential as effective biocontrol agents against *R. solani* according to *in vitro* investigations (Islam *et al.* 2016). Their abilities to produce extracellular hydrolytic enzymes, including chitinase, β -1,3-glucanase and proteinase were also confirmed. The present study aimed to evaluate the effectiveness of these native *Trichoderma* isolates against *R. solani* on tomato seedlings under greenhouse conditions after seed treatments.

MATERIALS AND METHODS

Fungal strains

Three strains (*T. harzianum* TR05, *T. virens* TR06 and *T. asperellum* TR08) isolated from various regions of Bangladesh during 2012–2013 (Islam *et al.* 2016) were used in this study. As a pathogen, *Rhizoctonia solani* MAFF241953 obtained from the National Institute of Agrobiological Sciences, Japan, was used after its pathogenicity was confirmed by artificial inoculations on tomato seedlings.

Islam et al.

Seed treatment

The effectiveness of the three Trichoderma strains against R. solani MAFF241953 on tomato seedlings was examined in a greenhouse following seed treatments. Inoculation of the Trichoderma strains and R. solani MAFF241953 were conducted according to the following combinations: $SE_0 = Control$, $SE_1 = TR05 +$ pathogen, $SE_2 = TR06 + pathogen$, $SE_3 = TR08 + pathogen$ and $SE_4 = only pathogen$. There were three replications per treatment.

First, a 12-d-old PDA-grown culture of each isolate was blended with sterile deionized water and a 30-ml fungal suspension was prepared. Spore density in the suspension was 7×10^8 spore/ml, determined by a haemocytometer under a light microscope. Tomato seeds were sterilized in a 1% sodium hypochlorite solution for 3 min and rinsed thoroughly in sterile distilled water. Inoculation with the Trichoderma strain was then performed by dipping seeds in the fungal suspension for 30 min. Control seeds were soaked in an equal volume of deionized water. The treated and control seeds were directly sown into trays $(12' \times 8'' \times 3'')$ filled with autoclaved commercial culture soil (0.8 kg/tray) at the rate of 50 seeds per tray.

R. solani MAFF241953 established on corn meal in a conical flask was applied to the trays at the rate of 5 g/kgsoil at 7 d before seed sowing. Trays were placed on a bench in a greenhouse. For 2 weeks after sowing, seedling emergence was monitored to determine effectiveness of the Trichoderma treatments on germination.

The percent disease incidence was determined 4 weeks after sowing using the following formula: Disease incidence (%) = $\frac{\text{Number of infected plants}}{100} \times 100$. The tomato seedlings were then removed from each tray

Total number of plants

and the roots gently washed using tap water. The effects of Trichoderma isolates on the growth of tomato seedlings were evaluated and recorded as follows: shoot length, root length and dry weights measured after drying for 5 d at 45°C. Seedling vigour was calculated using the following formula: Vigour index = (Root length + Shoot length) \times Seed germination percentage.

Statistical analysis

Tukey's test was performed to determine statistical differences in antifungal activities among Trichoderma isolates using the statistical software KyPlot version 2.0 beta 15.

RESULTS AND DISCUSSION

Figure 1 shows the effects of Trichoderma isolates on disease incidence of R. solani MAFF241953, growth of tomato seedlings and seedling vigour in the seed treatment. TR05 (SE₁) showed statistically lower disease incidence (5.79%) than TR06 (SE₂: 35.7%) and TR08 (SE₃: 21.6%) (Fig. 1A). In general, seed coats bearing Trichoderma sp. inoculum can give systemic protection against many seed-borne diseases (Linda 2000). Trichoderma is also well known to provide plants with useful molecules such as glucose oxidase and growthstimulating compounds that can increase their vigour and as a result resistance to pathogens (Brunner et al. 2005). Additionally, Trichoderma can produce antibiotics such as gliotoxin, viridian, cell wall degrading enzymes and also biologically active and heat stable metabolites such as ethyl acetate. These substances are mainly known to be involved in suppression of disease incidence. Moreover, T. viride and T. harzianum were reported as the best antagonists against several soil and seed-borne plant pathogens (Poddar et al. 2004).

The highest germination percentage (97.3%) was for the control (SE₀) and the lowest (55.3%) was for SE₄ (only pathogen). TR05 (SE₁) showed statistically higher germination percentage (89.7%) than TR06 (SE₂: 81.3%) and TR08 (SE₃: 73.3%) (Fig. 1B). The highest root length (13.5 cm) was for SE₁ (TR05) and the lowest (2.9 cm) for SE_4 (only pathogen) which was statistically similar with control (SE_6 : 6.70 cm) (Fig. 1C). TR05 (SE_1) had statistically greater shoot length (12.7 cm) than TR06 (SE₂: 8.27 cm), TR08 (SE₃: 5.73 cm) and control (SE₆: 6.67 cm), and the lowest (3.00 cm) was for SE₄ (only pathogen) (Fig. 1D). The highest dry weight (5.40 g) was for TR05 (SE₁) followed by TR06 (SE₂: 3.00g), and the lowest (0.41g) for SE₄ (only pathogen) was not statistically different from both the control (SE₀: 2.0 g) and TR08 (SE3: 0.99 g) (Fig. 1E). In these study, the inoculation with TR05, TR06 and TR08 not only suppressed collar rot disease but also enhanced germination percentage, root and shoot growth, dry weight and vigour of tomato seedlings compared to the infected control. Among the three Trichoderma isolates, TR05 exhibited significant enhancement of the above mentioned plant growth characteristics after the seed treatment (Fig. 1). Some researchers have reported that Trichoderma increased plant growth and productivity (Harman 2006; Manju and Mall, 2008). In this study, isolate TR05 gave the highest germination percentages. These studies have been confirmed in the case of T. harzianum and T. virens enhancing seed germination, root and shoot length (Dubey et al. 2007) as well as increasing the frequency of healthy plants. Lo and Lin (2002) also screened different Trichoderma strains for effects on plant growth and root growth of bitter gourd, luffa and cucumber and noted that they significantly increased seedling height, root exploration, leaf area and dry weight at 15 d after sowing.

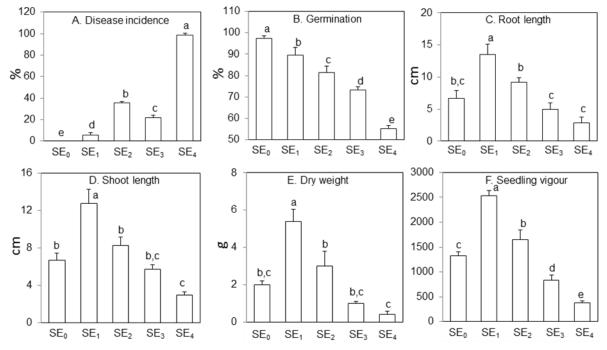


Fig. 1. Effects of the seed treatment with *Trichoderma* TR05, TR06 and TR08 on tomato seedlings infected by *R. solani* MAFF241953 (at four weeks): $SE_0 = \text{control}$, $SE_1 = \text{TR05} + \text{pathogen}$, $SE_2 = \text{TR06} + \text{pathogen}$, $SE_3 = \text{TR08} + \text{pathogen}$ and $SE_4 = \text{only pathogen}$. Bars indicate standard deviations (n=3). Different letters indicate significant differences according to Tukey's test (*P*<0.05).

TR05 (SE₁) had statistically higher seedling vigour index (2532.4) than TR06 (SE₂: 1647.5), TR08 (SE₃: 837.6) and control (SE₀: 1325.3), and the lowest of 376.6 was for SE₄ (only pathogen) (Fig. 1F). The vigour index showed similar trends to the germination percentages (Fig. 1). Mukhtar (2008) found that seed treatment with *T. harzianum* gave the highest germination index in okra and that *T. harzianum* could be useful to enhance the germination percentage as well as seedling vigour. Begum *et al.* (2010) observed that application of *T. harzianum* IMI 392432 significantly suppressed *Alternaria* fruit rot disease of chili and improved the seed germination percentage, vigour, growth and yield. Other investigators reported that seeds treated with *T. viride*, *T. harzianum* and *T. pseudokoningii* inoculant extracts showed increased seed germination rates and seedling vigour and reduced incidence of seed-borne fungal pathogens compared to controls (Zheng and Shetty, 2000; Bharath *et al.* 2006).

CONCLUSION

Results from the present investigation suggests that, applications of *T. harzianum* TR05 as a seed treatment has potential for controlling foot rot of tomato seedlings caused by *R. solani*. Further studies are underway to examine the effectiveness of these isolates as alternatives to chemicals to control this disease in different field conditions.

REFERENCES

Anahosur KH (2001) Integrated management of potato Sclerotium wilt caused by *Sclerotium rolfsii*. Indian *Phytopathol*. 54: 158-166.

Babar HM (1999) Studies on collar rot of sunflower. Ph.D thesis. Department of Plant Pathology. Bangladesh Agricultural University, Mymensingh. p.189.

Begum MF, Rahman MA, Alam MF (2010) Biological control of Alternaria fruit rot of chili by *Trichoderma* species under field conditions. *Mycobiol.* 38(2), 113-117.

Bharath BG, Lokesh S, Prakash HS, Shetty HS (2006) Evaluation of different plant protectants against seed mycoflora of watermelon (*Citrullus lanatus*). *Res J Bot*. 16:1-5.

Brunner K, Zeilinger S, Ciliento R, Woo SL, Lorito M, Kubicek CP, Robert LM (2005) Improvement of the fungal biocontrol agent *Trichoderma atroviride* to enhance both antagonism and induction of plant systemic disease resistance. *Appl Environ Microbiol*. 71(7), 3959-3965.

Dubey SC, Suresha M, Singha B (2007) Evaluation of *Trichoderma* species against *Fusarium oxysporum* f. sp. *ciceris* for integrated management of chickpea wilt. *Biol Cont.* 40: 118-127.

Islam et al.

Fakir GA (2000) List of seed-borne diseases of important crops occurring in Bangladesh. Seed Pathology Laboratory. Department of Plant Pathology. Bangladesh Agricultural University, Mymensingh. p.18.

Harman GE (2006) Overview of mechanisms and uses of Trichoderma spp. Phytopathol. 96: 190-194.

Islam MM, Hossain DM, Rahman MME, Suzuki K, Narisawa T, Hossain I, Meah MB, Nonaka M, Harada N (2016) Native *Trichoderma* strains isolated from Bangladesh with broad spectrum of antifungal action against fungal phytopathogens. *Arch. Phytopathol.* Plant Protect. 49: 75-93.

Linda EH (2000) Reduction of Verticillium wilts symptoms in cotton following seed treatment with *Trichoderma virens. J Cotton Sci.* 4: 224-231.

Lo CT, Lin CY (2002) Screening strains of *Trichoderma* spp. for plant growth enhancement in Taiwan. Plant Pathology Bulletin. 11: 215-220.

Manju S, Mall TP (2008) Efficacy of *Trichoderma* species on *Phytopthora dresceleri* f.sp. *cajani* of Pigeon pea. *Ann Plant Prot Sci.* 16: 162-164.

Mukhtar I (2008) Influence of Trichoderma species on seed germination in okra. Mycopath. 6: 47-50.

Naseby DC, Pascual JA, Lync JM (2000) Effect of biocontrol strains of *Trichoderma* on plant growth, *Pythium ultimum* population, soil microbial communities and soil enzyme activities. *J Appl Microbiol.* 88: 161-169.

Poddar RK, Singh DV, Dubey SC (2004) Integrated application of *Trichoderma harzianum* mutants and carbendazim to manage chickpea wilt (*Fusarium oxysporum* f.sp. *ciceri*). *Indian J Agric Sci*. 74: 346-348.

Warrior P, Kondru K, Preeti V, Vasudevan P (2002) Formulation of biocontrol agents for pest and disease management. In: *Biological Control of Crop Disease*. Gnanamanickam SS. (ed.). Marcel Dekker, New York.

Zheng Z, Shetty K (2000) Enhancement of pea (*Pisum sativum*) seedling vigour and associated phenolic content by extracts of apple pomace fermented with *Trichoderma* spp. *Process Biochem.* 36: 79-84.